

Targeted Therapy in Lung Cancer

Colin Hardin¹ and Haiying Cheng^{1*}

¹Department of Oncology, Montefiore Medical Center/Albert Einstein College of Medicine, USA

***Corresponding author:** Haiying Cheng, Department of Oncology, Montefiore Medical Center/Albert Einstein College of Medicine, USA, Tel: 718-430-2430; Email: hcheng@montefiore.org

Published Date: August 07, 2016

INTRODUCTION TO TARGETED THERAPY IN LUNG CANCER

Advances in understanding the molecular mechanisms of lung cancer oncogenesis and progression have shifted the treatment paradigm for this deadly disease. Targeting the “driver” genomic alterations including Epidermal Growth Factor Receptor (**EGFR**) mutations, Anaplastic Lymphoma Kinase (**ALK**) rearrangements, and ROS1 translocations has led to improved clinical outcomes in subgroups of patients with lung adenocarcinoma.

Decoding of the human cancer genome and identification of oncogenic driver mutations have been important milestones for the treatment of Non-Small Cell Lung Cancer (**NSCLC**). The Cancer Genome Atlas (**TCGA**) Research Network has successfully profiled lung adenocarcinoma and squamous cell lung cancers in an effort to characterize driver mutations [1,2]. Most oncogenic driver mutations are found in adenocarcinoma, where over 50% of tumors harbor genomic aberrations [1,3,5]. Characterization of genomic alterations in squamous cell lung carcinoma has also shed light on “personalized therapy” although targetable drivers have are less frequently observed [2,3].

In this chapter, we will review the biology, highlight existing therapy, and discuss emerging treatment strategies for distinct molecular subtypes of NSCLC.

EGFR-TARGETED THERAPY

The EGFR gene encodes for a transmembrane tyrosine kinase receptor. Mutations can lead to aberrant EGFR signaling, tumor growth and cellular proliferation [6]. The majority of patients (85-90%) with EGFR mutations harbor in-frame deletions in exon 19 or L858R in exon 18. Activating EGFR mutations such as deletion 19, exon 21 (L858R, L861Q), exon 18 (G719X), and exon 20 (S768I) are associated with sensitivity to EGFR Tyrosine Kinase Inhibitors (**TKIs**) [6-11], while exon 20 insertion mutations confer resistance. EGFR mutations are more common in women, Asians, and those with no or light history of smoking [9,12]. There are four FDA approved, orally bioavailable, EGFR TKIs in the United States: the first generations of erlotinib (Tarceva, OSI Pharmaceuticals) and gefitinib (Iressa, Astra-Zeneca), the second generation of afatinib (Gilotrif, BoehringerIngelheim), and the third generation of osimertinib (Tagrisso, Astra-Zeneca) [13].

First Generation EGFR TKI: Erlotinib and Gefitinib

The BR.21 trial evaluated erlotinib versus placebo in the second or third line setting and demonstrated a benefit in Overall Survival (**OS**) in patients with stage IIIB or IV NSCLC (6.7 versus 4.7 months, HR 0.70, $p < 0.001$) [14], leading to the approval of erlotinib for advanced NSCLC.

Later studies indicated that the presence of EGFR mutations is predictive of response to targeted therapy with EGFR TKIs, such as erlotinib [12]. First line TKI monotherapy in EGFR mutant lung cancer demonstrated improved Progression Free Survival (**PFS**) compared to chemotherapy [15-18]. The Iressa Pan-Asia Study (**IPASS**) phase III trial compared untreated patients in East Asia treated with gefitinib or carboplatin plus paclitaxel. Gefitinib was associated with superior 12 month PFS: 24.9% versus 6.7% (HR 0.74; 95% CI: 0.65-0.85; $P < 0.001$). Furthermore, subgroup analysis predicted better outcomes in patients with EGFR mutations [15]. The phase III WJTOG3405 trial compared chemotherapy naïve Japanese patients with advanced EGFR mutant NSCLC treated with gefitinib versus cisplatin plus docetaxel. Gefitinib prolonged PFS to 9.2 months (95% CI: 8.0-13.9) compared to 6.3 months (95% CI: 5.8-7.8) [16]. The OPTIMAL trial evaluated first line platinum doublet therapy compared to erlotinib in EGFR mutant patients with advanced NSCLC. Erlotinib was associated with median PFS of 13.1 months versus 4.6 months with chemotherapy, while activating EGFR mutations were the most important factor predicting treatment efficacy [18]. The EURTAC trial was the first randomized trial targeting a non-Asian population in EGFR positive NSCLC patients. It compared erlotinib to standard platinum doublet as first line therapy; median PFS was 9.7 months (95% CI: 8.4-12.3) in the erlotinib group, versus 5.2 months (95% CI: 4.5-5.8) with chemotherapy (HR 0.37, 95% CI: 0.25-0.54; $p < 0.0001$) [19].

On the other hand, the phase III TRIBUTE study compared untreated NSCLC patients treated with carboplatin and paclitaxel combined with either erlotinib or placebo. No improvement in OS was gained by adding erlotinib to chemotherapy, but subgroup analysis of those patients with EGFR mutant or KRAS wild type tumors found both were associated with erlotinib response [20,21].

Finally, the SATURN trial demonstrated that erlotinib maintenance led to both prolonged PFS and OS in patients who had stable disease after traditional chemotherapy (HR 0.72; $p=0.0019$). Interestingly, the erlotinib-related OS benefit was significant in all subgroups irrespective of EGFR mutation status [22].

Second Generation EGFR TKI: Afatinib

Afatinib binds irreversibly to wild-type and aberrant EGFR. It also inhibits ErbB2, ErbB4, ErbB family dimers and the transphosphorylation of ErbB3 [23].

LUX-Lung 3, a randomized phase III study of treatment naïve patients with stage IIIB/IV adenocarcinoma, compared afatinib to cisplatin/pemetrexed (**CP**). Median PFS was 11.1 months with afatinib compared with 6.9 months for patients treated with CP (HR 0.58; 95% CI: 0.43-0.78; $p=0.001$). In patients with exon 19 deletions and L858R EGFR mutations, PFS benefit was even larger (13.6 months versus 6.9 months). This trial is also notable for enrolling a large non-Asian population [24,25]. The LUX-Lung 6 trial compared afatinib to gemcitabine/cisplatin in first line treatment of an Asian population with EGFR mutant advanced NSCLC. PFS was significantly extended in the afatinib group (11.0 months) compared to the gemcitabine/cisplatin group (5.6 months) [26].

Both LUX-Lung 3 and 6 identified improved PFS with afatinib based on EGFR mutation type; PFS was most improved in those individuals with exon 19 deletion [24,26]. Neither trial identified improved OS but preplanned subgroup analysis demonstrated those patients with EGFR mutations had improved OS with afatinib. In LUX-Lung 3, OS was 33.3 months in the afatinib group compared to 21.1 months with chemotherapy (HR 0.54, 95% CI: 0.36-0.79, $p=0.0015$); in LUX-Lung 6, OS was 31.4 months versus 18.4 months (HR 0.64, 95% CI: 0.44-0.94, $p=0.023$) [27].

The phase IIb LUX-Lung 7 trial compared afatinib to gefitinib as first line therapy in EGFR-mutant NSCLC. At median follow up of 27.3 months, PFS was 11.0 months with afatinib compared to 10.9 months with gefitinib (HR 0.73, 95% CI: 0.57-0.95, $p=0.017$). OS data was not mature at the time of publication. Despite preclinical data that afatinib could overcome T790M-mediated resistance, this has not been observed in patients likely due to failure to reach preclinical concentrations *in vivo* as a result of dose-limiting toxicity [28].

Third Generation EGFR TKI: Osimertinib and Rociletinib

The initial response to EGFR TKIs and prolongation of PFS is often impressive, but the majority of patients will progress after 1 year [15,19,24,29]. Approximately 60% of acquired EGFR resistance results from exon 20 T790M mutations [30-32]. Alternative bypass pathways also contribute, including PI3K/AKT, MAP kinase, JAK/STAT, BRAF mutation, and ERBB2/MET amplification [4,32-34]. Histological transformation is the third mechanism of acquired EGFR resistance, via conversion to small cell histology [35] or epithelial to mesenchymal transition [36]. Due to the variability and frequency of secondary mutations after initial therapy, repeat biopsy after progression on first line EGFR TKI is becoming the standard of care [4].

Osimertinib (AZD9291, AstraZeneca) is a third generation EGFR TKI approved for patients with EGFR T790M after progression on prior EGFR TKI treatment [37]. Osimertinib has nearly 200 times higher potency to EGFR harboring L828R/T790M mutations compared to wild-type [38]. The phase I/II AURA trial evaluated patients treated with osimertinib who had progressed after prior EGFR TKI treatment: Overall Response Rate (**ORR**) was 61% (95% CI: 52-70) with EGFR T790M compared to 21% (95% CI: 12-34) in those without T790M [39]. PFS in EGFR T790M-positive patients was 9.6 months (95% CI: 8.3 - not reached) compared to 2.8 months (95% CI: 2.1-4.3). Adverse events included diarrhea, rash, nausea, and decreased appetite. Osimertinib has also induced central nervous system responses, an observation not made with earlier generations of EGFR TKIs [39]. In the BLOOM phase I study, preliminary data from 21 patients with leptomeningeal disease showed intracranial radiological and neurologic responses in seven and five patients, respectively [40].

Rociletinib (CO-1686) is another oral, irreversible, potent inhibitor of activating EGFR mutations as well as the T790M mutation [41]. Rociletinib was evaluated in the TIGER-X phase I/II trial in patients with T790M-positive tumors. Similar adverse events were noted to osimertinib, but additional toxicities including QTc interval prolongation (20.6%), hyperglycemia (46.9%), and cataracts (53%) were observed [42-44]. Initial data was promising [42,43] but updated results presented at ASCO 2016 noted more modest ORR (21.8%) and duration of response (12.2 months) [45].

Novel EGFR TKIs continue to emerge. For instance, AZD3759 was also evaluated in the BLOOM trial, where encouraging intracranial anti-tumor activity was observed and toxicities were limited to diarrhea and skin rash [46]. Expansion cohorts are ongoing (NCT02228369).

Future Directions– Plasma Based Mutation Testing

The cobas EGFR Mutation Test v2 (Roche) is the first peripheral blood-based liquid biopsy test approved by the FDA. It was designed for evaluation of EGFR mutations. Detection of circulating free tumor DNA (ctDNA) or circulating tumor cells is an emerging strategy for tumor genotyping due to the importance of tumor genetics in treatment selection. The development of non-invasive genotyping allows for rapid detection of EGFR mutations and continual monitoring for additional resistance mechanisms, specifically T790M [47]. Existing techniques for tumor biopsy include fine needle aspirations and image-guided or endoscopic biopsies that place a burden on patients at the time of progression and carry additional risk. Plasma-based assays provide for rapid and non-invasive monitoring in what is now established as a molecularly dynamic disease. Next generation sequencing and digital PCR have allowed for detection of ctDNA at levels as low as 0.1% of total blood DNA [48]. Positive predictive value appears to approach that of tissue-based testing, while the presence of the T790M EGFR mutation has 81% positive concordance with tissue testing [49]. Limitations exist because not all tumors “shed” ctDNA, and the complexity of tumor heterogeneity cannot yet be replicated by plasma testing. Therefore, the two techniques currently appear best suited for a complimentary approach.

Uncommon EGFR Mutations

Approximately 10% of EGFR mutations consist of so called “uncommon” mutations, including G719X, L861Q, S768I, G719X + L861Q, and G719X + S768I. Patients with these mutations display inferior response rates (41.6% versus 66.5%; $p < 0.001$), PFS (7.7 versus 11.4 months; $p < 0.001$) and OS (24.0 versus 29.7 months; $p = 0.005$) compared to individuals with common EGFR mutations [50]. Moreover, the exon 20ins mutation is associated with particularly poor outcomes (ORR 8.3%) [51]. Additional studies to evaluate the efficacy of second and third generation EGFR TKIs in this EGFR subgroup are ongoing.

ANAPLASTIC LYMPHOMA KINASE (ALK)

The ALK gene encodes a receptor tyrosine kinase not expressed in normal lung tissue [52]. ALK is more typically expressed during embryogenesis and neuron development [53,54]. In lung cancer, oncogenic fusion genes comprising the echinoderm microtubule-associated protein-like 4 (**EML4**) and ALK genes have been observed in 2 to 7% of NSCLC [55-57]. Examples of more rare alternative fusion partners with ALK have also been identified [58,59]. EML4-ALK fusion produces a gain of function mutation that results in constitutive activation [55,58,60,61]. ALK fusions are mutually exclusive of KRAS and EGFR mutations, but occur in patients with similar characteristics (ie. adenocarcinoma, light/never smokers, and young patients) [56,58,62-65].

First Generation ALK TKI: Crizotinib

Crizotinib (Xalkori, Pfizer) is an oral ATP-competitive selective inhibitor of ALK, ROS-1, and MET tyrosine kinases [66-68] and is FDA approved for treatment of ALK-positive NSCLC. PROFILE 1007 was the first phase III trial comparing crizotinib to chemotherapy in patients with locally advanced or metastatic ALK-positive lung cancer as second line therapy [69]. PFS was 7.7 months in the crizotinib group compared to 3.0 months with pemetrexed or docetaxel (HR 0.49; 95% CI: 0.37-0.64). There was no significant improvement in OS. The main adverse effects of crizotinib included visual disorders, gastrointestinal side effects and elevated liver transaminases. PROFILE 1014 evaluated crizotinib versus chemotherapy as first line treatment for advanced ALK translocation-positive NSCLC. Crizotinib significantly improved PFS (10.9 versus 7.0 months; HR 0.45, 95% CI: 0.35-0.60) and ORR (74% versus 45%, $p < 0.001$) [70].

Second Generation ALK TKI: Ceritinib and Alectinib

Unfortunately, acquired resistance to crizotinib generally occurs within one year. In 30% of patients resistance is due to secondary ALK mutations including L1196M, F1174L, S1206Y, G1269A, and G1202R [71,72]. ALK resistance can also be due to ALK amplification (6-16%), increased EGFR signaling (30-35%), and change in driver mutation (10-15%) [73,74]. A common site of progression is to the CNS: by 2 years, 45.5% of patients will have brain metastatic disease [75] due to low CNS penetration by crizotinib [76,77].

Ceritinib (LDK378; Zydakia; Novartis) is a second generation ALK TKI with greater potency compared to crizotinib that inhibits some of the secondary ALK mutations that cause crizotinib resistance [71,78-80]. Ceritinib also inhibits IGF-1 and ROS1, but not MET. Ceritinib was granted accelerated approval in 2014 based on the ASCEND-1 trial, an open-label, phase 1 trial evaluating ceritinib in patients with ALK-rearranged locally advanced or metastatic cancer that had progressed despite prior ALK inhibitor treatment. ORR was 44% (95% CI: 36-52) combined with a manageable side effect profile (diarrhea, nausea, vomiting and increased transaminases) [81,82]. Ceritinib has increased bioavailability in the CNS, as evidenced by observed responses in CNS disease [83].

Alectinib (AF-001JP; Alecensa; Chugai Pharmaceuticals) is a highly selective ALK inhibitor with activity against wild-type ALK and mutated ALK, including the L1196M gatekeeper mutation. It also has activity against RET and IGF-1 [84,85]. It was granted accelerated approval for treatment of crizotinib resistant, ALK positive, metastatic NSCLC. A Japanese phase I/II trial evaluated 46 ALK-inhibitor naïve patients treated with alectinib: ORR was 93.5% with mean follow up of 14.8 months. The most common adverse events were dysgeusia, elevated liver function tests and creatinine, constipation and rash [86]. Phase I data from a North American trial evaluating alectinib after progression on crizotinib confirmed tolerability with the most common adverse events being fatigue, myalgias and edema [87]. Phase II data published in 2016 found 33 of 69 patients had a response to alectinib (ORR 48%, 95% CI: 36-60) [88]. Of this group, 52 patients had CNS disease at baseline; median duration of response to alectinib was 11.1 months. 46 (89%) patients achieved disease control (95% CI: 77-96), including 13 (25%) with a complete response [88]. A second, global, phase II study evaluating alectinib in previously treated patients with advanced ALK positive NSCLC reported similar results including ORR of 50% with PFS of 8.9 months. CNS response was also noted: in 84 patients with CNS metastases, 23 (27%) had observed CNS CR, and overall disease control rate was 83% (95% CI: 74-91) [89]. J-ALEX is a Japanese phase III trial comparing first line alectinib to crizotinib in ALK positive advanced NSCLC that included patients with measurable brain metastases. First interim analysis data found median PFS in the crizotinib group was 10.2 months (95% CI: 8.2-12), and not reached with alectinib (95% CI: 20.3-NR) (HR 0.34, 95% CI: 0.17-0.71, $p < 0.0001$). Subgroup analysis of PFS in patients with brain metastases found statistically superior outcomes for patients treated with alectinib [90]. ALEX is the North American counterpart to this trial and has completed recruitment (NCT02075840).

Third Generation ALK TKI: Lorlatinib and Brigatinib

Lorlatinib (PF-06463922, Pfizer) is a reversible, third generation, ALK inhibitor with improved brain bioavailability and broad-spectrum potency against ALK-resistant mutations [91]. In a phase I clinical trial evaluating its use in patients resistant to both crizotinib and ceritinib, patients with history of one or more prior ALK TKI therapies had an ORR of 46% (95% CI: 31-63%) and PFS of 11.4 months (95% CI: 3.4-16.6 months) [92]. The most common toxicities noted were hypercholesterolemia, peripheral neuropathy and edema [93].

Lorlatinib treatment may result in acquisition of the L1198F mutation, which paradoxically restores crizotinib sensitivity [94]. Lorlatinib is the first ALK TKI with identified activity against the G1202R mutation [91].

Brigatinib (AP26113) is a next generation ALK inhibitor with high selectivity towards clinically relevant ALK mutations, including L1196M, and activating EGFR mutations, including T790M [95,96]. Breakthrough therapy designation was granted in 2014. Phase I/II data has been reported by Rosell et al. (2016); 79 patients with ALK positive NSCLC (70 with prior treatment with crizotinib) treated with brigatinib had an ORR of 71% and median PFS of 13.4 months. 53% (8/15) of patients with measurable CNS disease had an intracranial objective response, and median duration of response in these patients was 18.9 months. The most common adverse effects were nausea, fatigue, diarrhea, hypertension and cough [97,98]. ALTA, a phase II trial of brigatinib in patients with ALK positive NSCLC who progressed on crizotinib, has completed recruitment of patients (NCT02094573). Preliminary data included 67% with CNS disease and 74% with prior treatment who were treated at 180mg daily. Reported PFS 12.9 months (95% CI: 11.1-NR), ORR 54% (97.5% CI: 43-65) and CNS response 67% compare favorably with previously reported outcomes with ceritinib and alectinib [81,89].

ROS1-TARGETED THERAPY

ROS1 is related to ALK and is normally expressed in tissues during development with little to no expression in normal adult tissues [99,100]. ROS1 fusion results in oncogenic activation. Rearrangements occur in only 1-2% of patients with NSCLC [101]. Those patients identified with ROS1 rearrangements share similar characteristics to those with EGFR mutations, including younger age, never-smokers, and Asian ethnicity [102]. ROS1 mutations are mutually exclusive to both EGFR and ALK [56,101-103].

In an expanded phase I study evaluating crizotinib in patients with NSCLC who had tested positive for ROS1, ORR was 72% (95% CI: 58-84), median duration of response was 17.6 months (95% CI: 14.5 to not reached) and PFS was 19.2 months (95% CI: 14.4 to not reached) [104]. Toxicities in patients with ROS1 were similar to that in patients treated with ALK mutations [104]. Despite dramatic improvements in PFS compared to historical means, eventual resistance to ROS1 inhibition usually develops [105].

Second Generation ROS1 Inhibition

In ROS1 NSCLC, the CNS is a common site of relapse while acquired mutations, including G2032R and D2033N, that confer TKI resistance have been identified [105,106]. Lorlatinib, the CNS penetrant, dual ALK/ROS1 TKI, has demonstrated activity in ROS1 mutant patients with acquired resistance to crizotinib, including those with CNS disease [92,107]. Described toxicities include hyperlipidemia, peripheral edema, and rarely, neurocognitive disturbance. The cMET/RET/VEGFR inhibitor cabozantinib (XL184) has demonstrated activity against crizotinib resistant CD74-ROS1 fusion [108], and in one patient treatment led to rapid clinical and radiographic response [106].

KRAS

KRAS is the most commonly altered gene in NSCLC, comprising 25 to 32% of mutations [1,109,110]. Despite extensive study KRAS remains a poor target for therapy. Downstream effectors of KRAS have been evaluated and show more promise, particularly MEK1/2 and BRAF. However, a growing understanding of KRAS biology and its downstream pathways has led to renewed optimism and new clinical studies evaluating KRAS inhibition [111].

KRAS mutations are strongly associated with smoking [112]. There is also data that KRAS mutations may impart a worse prognosis compared to wild type [113-115]. KRAS mutation may be a negative predictive factor of EGFR TKI response, but no consensus regarding the clinical value of KRAS mutations in NSCLC exists [116,117].

Downstream KRAS Inhibitors: MEK Inhibitors

MEK1 and MEK2 are downstream effectors of KRAS; two MEK1/2 inhibitors, selumetinib and trametinib have been studied in NSCLC [118]. A prospective phase II trial combining docetaxel with and without selumetinib in patients with KRAS mutant NSCLC who progressed after first line chemotherapy did not demonstrate significantly improved OS, but the combination did lead to statistically significant improvements in PFS and patient reported outcomes. The main non-hematologic side effects reported were rash, diarrhea, fatigue and edema [119]. Combining selumetinib with erlotinib did not show improved ORR or PFS compared to erlotinib alone in either KRAS mutant or wild type patients [120].

In phase I evaluation studies, trametinib has exhibited a side effect profile that includes rash, diarrhea, and CNS retinopathy [121]. Synergy with chemotherapy has been evaluated and shows modest promise in early stage trials [118,122,123].

BRAF V600E

BRAF is a serine-threonine kinase that belongs to the RAF kinase family and is downstream of KRAS [124]. Frequency of BRAF mutations in NSCLC is approximately 1-5%, but is more common in those with a history of smoking [1,3]. V600E is the most commonly observed mutation in BRAF NSCLC [125] and is associated with more aggressive disease and worse OS [126]. Initial case reports documented activity of BRAF inhibitors in V600E positive NSCLC [127,128], including one report of CNS response [129]. Later phase II data in dabrafenib treated BRAF V600E mutated NSCLC has shown ORR of 54%. Adverse effects were decreased appetite, fatigue, asthenia, dyspnea, and nausea [130]. BRAF V600 positive NSCLC patients have also been evaluated in a phase II “basket study” with ORR of 43% (95% CI: 18-71) [131]. The MEK inhibitor trametinib has also been combined with dabrafenib, increasing ORR to 63% (95% CI: 40-81.2) [132]. Despite no FDA approval for BRAF inhibitors in NSCLC, current NCCN guidelines support the use of vemurafenib and dabrafenib alone or in conjunction with trametinib in patients with BRAF V600E mutations [37].

RET

RET acts as a proto-oncogene in a small subset of patients with NSCLC, primarily those with adenocarcinoma [1,3,133,134]. RET rearrangements result in gain of function mutations. Four RET fusion partners have been identified in NSCLC, including CCDC6-RET and KIF5B [134-136]. KIF5B-RET can be inhibited via TKIs with anti-RET activity including sunitinib, sorafenib, vandetanib and lenvatinib [137-140]. Clinical trials evaluating the addition of anti-RET therapy to standard chemotherapy in non-RET selected NSCLC have demonstrated limited improvement in ORR [141-143]. However, RET-fusion positive NSCLC appears to respond well to cabozantinib [136]. Vandetanib is also active in RET selected patients [144,145]. Phase II Japanese data evaluating vandetanib in advanced stage RET fusion positive non-squamous NSCLC described ORR of 20% (95% CI: 3-56) in KIF5B-RET and 83% (95% CI: 36-99.6) in those with CCDC6-RET [146]. The most common adverse events were hypertension, diarrhea, and rash. Analysis of a global registry of RET-rearranged NSCLC has identified off-label activity using cabozantinib, vandetanib, sunitinib, sorafenib, alectinib, lenvatinib, nintedanib and ponatinib [146].

ERBB2/HER2

Human epidermal growth factor 2 (HER2 or ERBB2) mutations are oncogenic drivers in approximately 2% of NSCLC cases [1,147]. HER2 mutations are most commonly identified in non-smokers, females, and those with adenocarcinoma [148]. The most common mutations are in-frame insertions in exon 20, leading to uncontrolled activation of downstream AKT and MEK [149]. In addition, up to 20% of patients with NSCLC have HER2 amplification or HER2 protein over expression [150-152]. The addition of trastuzumab to standard therapy in HER2-amplified NSCLC has not shown significant benefit [153-155].

Afatinib has activity against EGFR and HER2 [23], and a small phase II trial evaluated afatinib in HER2 mutation positive NSCLC. Partial response or stable disease was noted in 3 of 5 patients, although dose limiting toxicity likely limited benefit as higher doses are required compared to those used in EGFR mutant NSCLC [156]. The pan-HER TKI dacomitinib irreversibly binds to HER2, HER1, and HER4 tyrosine kinases. NSCLC patients with HER2 mutations and amplifications treated with dacomitinib were studied in a phase II trial: 3 of 26 patients with HER2 exon 20 mutation positive tumors had partial responses, while 0 of 4 patients with HER2 amplification had documented response [157].

MET

MET, or Hepatocyte Growth Factor Receptor (**HGFR**), is a receptor tyrosine kinase that activates the PI3K/AKT and MAPK pathways [158]. MET over expression has been identified in approximately 20% of EGFR mutant NSCLC cases with acquired resistance to EGFR TKIs [159,160]. Over expression is associated with a poor prognosis [161]. In comparison, aberrant MET expression acts as a driver mutation in only 2% of lung adenocarcinoma [1,162,163].

Crizotinib has activity against MET and has demonstrated anti-tumor activity in patients with MET amplifications [164,165]. Several anti-MET therapies including tivantinib, crizotinib, and cabozantinib have been tested in advanced NSCLC [166-169]. Rilotumumab, a fully humanized IgG2 monoclonal antibody that blocks HGF, is being tested in combination with erlotinib in patients with recurrent or advanced NSCLC (NCT01233687).

MET exon 14 skipping mutations occur in 3-4% of NSCLC, and are more common in pulmonary sarcomatoid carcinoma [1,160,163,170]. Patients with MET exon 14 splice variants have demonstrated responses to MET-directed therapy, including crizotinib [171].

PTEN/PI3K/AKT/MTOR

The PI3K/AKT/mTOR pathway plays an important role in cell growth and survival and is negatively regulated by PTEN [172]. Ubiquitous activation can lead to inhibition of pro-apoptotic and regulatory functions [173,174]. PTEN inactivating mutations occur in 4-5% of NSCLC and an additional 70% of NSCLC are associated with reductions or loss of PTEN expression. Meanwhile, mutations in PI3K and AKT occur in 2-5% and 1-2%, respectively [175,176]. PI3K mutations commonly coexist with EGFR and KRAS mutations [177]. Increased AKT activity can lead to acquired resistance of EGFR-targeted therapy [178,179]. Mutations in the PI3K/AKT/mTOR pathway appear to be associated with a poor prognosis [175,176].

GDC 0941, a dual PI3K/mTOR inhibitor, has demonstrated encouraging activity in phase I evaluation when combined with MEK inhibition or carboplatin/paclitaxel [182,183]. The AKT inhibitor MK-2206 is being evaluated in a phase II trial (NCT01294306). XL-765 is an oral dual PI3K/mTOR inhibitor with phase I activity being evaluated in combination with erlotinib in advanced NSCLC (NCT00777699). The mTOR inhibitors everolimus and temsirolimus are FDA approved in several malignancies but there is no current FDA approval for mTOR inhibitors in NSCLC. Temsirolimus has been evaluated in phase II study: adverse events were tolerable but the study did not meet its primary objective [182]. Unselected populations with NSCLC treated with everolimus in combination with erlotinib [183], carboplatin/paclitaxel [184], or docetaxel [185] have reported discouraging results. In unselected populations of NSCLC, dual mTOR1/2 inhibitors have demonstrated limited clinical benefit [186], however, a recently published study detailing RICTOR amplification in NSCLC described a robust response observed with CC-223 and MLN0128, two dual mTORC1/2 inhibitors. RICTOR is a component of mTOR2; amplification occurs in between 8-13% of patients with NSCLC and this finding may suggest a role for genomic stratification specific to RICTOR amplification in the future [187].

EMERGING TARGETED THERAPY IN SQUAMOUS CELL LUNG CANCER

Squamous Cell Lung Cancer (**SQCL**) is strongly associated with tobacco use, and the subsequent genomic alterations observed are complex and highly variable [2]. Unfortunately, recurrent alterations in previously identified kinase genes occur much less frequently as compared to

adenocarcinoma. The most common variants in SQLC are loss of TP53 and CDKN2A, two targets that have historically been poor candidates for targeted therapy [188,189]. Additional alterations or aberrant genetic pathways have been identified in SQLC and include RAS (loss of NF1, RASA1, RAS), PI3K (PIK3A, PTEN, PIK3R1), oxidative stress regulation mutations (ie. NFE2L2/KEAP1/CUL3), differentiation pathways (NOTCH1, TP63 and SOX2 gain), and immune evasion (HLA and B2M) mutations [2,188,189]. RICTOR amplification has been identified in 7.4-15.8% of SQLC patients, and may be an emerging target for novel therapies [186,187].

EGFR over expression and amplification may rarely occur in SQLC. Evaluation of erlotinib in patients with previously treated SQLC found improved survival compared to placebo, resulting in its approval for second line and maintenance therapy [190,191]. Afatinib was evaluated in the second line setting versus erlotinib in the LUX-lung 8 trial. This resulted in a modest improvement of PFS (2.6 versus 1.9 months, HR 0.81, $p=0.0103$) and OS (7.9 versus 6.8 months, HR 0.81, $p=0.0077$) [192]. Necitumumab is a fully human IgG1 monoclonal antibody designed to target EGFR, inhibit DNA repair, and induce antibody dependent cell-mediated cytotoxicity [193]. The SQUIRE study compared necitumumab combined with gemcitabine and cisplatin versus chemotherapy alone as first line treatment in advanced NSCLC. Combination therapy attained statistical significance (OS 11.5 versus 9.9 months, HR 0.84; $p=0.01$) while modest improvement in PFS was also observed (5.7 versus 5.5 months, HR 0.85; $p=0.02$) [194]. Adverse effects were consistent with previous complications of EGFR inhibition including skin rash, hypomagnesemia, conjunctivitis and diarrhea, as well as increased rates of thromboembolic events [193,194].

CONCLUSION

Molecular targeted therapy plays an important role for many patients in NSCLC. FDA approved treatment in EGFR mutant NSCLC includes, erlotinib, gefitinib, and afatinib, while specific resistance via T790M can be targeted by osimertinib. ALK inhibitors including crizotinib, ceritinib and alectinib are approved, where as third generation inhibitors lorlatinib and brigatinib show promising activity. Crizotinib is approved to target ROS1, while lorlatinib and cabozantinib have shown clinical benefit. Ongoing study in NSCLC subsets including HER2/ERBB2, MET, KRAS, RET, NTRK and PI3K/PTEN/mTOR are underway. Taken together, advances in targeted therapy may further lead to shifting paradigms in the years to come. In addition, as the first plasma based ctDNA test was recently granted FDA approval for EGFR mutations, novel non-invasive biomarker tests also carry great promise.

References

1. Collisson EA, Campbell JD, Brooks AN, Berger AH, Lee W, et al. Comprehensive molecular profiling of lung adenocarcinoma. *Nature*. 2014; 511: 543550.
2. Hammerman PS, Lawrence MS, Voet D, Jing R, Jing R, et al. Comprehensive genomic characterization of squamous cell lung cancers. *Nature*. 2012; 489: 519-525.
3. Wang R, Zhang Y, Pan Y, Li Y, Hu H, et al. Comprehensive investigation of oncogenic driver mutations in Chinese non-small cell lung cancer patients. *Oncotarget*. 2015; 6: 1-9.

4. Shea M, Costa DB, Rangachari D. Management of advanced non-small cell lung cancers with known mutations or rearrangements: latest evidence and treatment approaches. *Ther Adv Respir Dis.* 2016; 10: 113-129.
5. Tsao AS, Scagliotti GV, Bunn PA, Carbone DP, Warren GW, et al. Scientific Advances in Lung Cancer 2015. *J Thorac Oncol.* 2016; 11: 613-638.
6. Han SW. Predictive and Prognostic Impact of Epidermal Growth Factor Receptor Mutation in Non-Small-Cell Lung Cancer Patients Treated With Gefitinib. *Journal of Clinical Oncology.* 2005; 23: 2493-2501.
7. Mitsudomi T, Kosaka T, Endoh H, Horio Y, Hida T, et al. Mutations of the epidermal growth factor receptor gene predict prolonged survival after gefitinib treatment in patients with non-small-cell lung cancer with postoperative recurrence. *Journal of Clinical Oncology.* 2005; 23: 2513-2520.
8. Bell DW, Lynch TJ, Haserlat SM, Harris PL, Okimoto RA, et al. Epidermal growth factor receptor mutations and gene amplification in non-small-cell lung cancer: Molecular analysis of the IDEAL/INTACT gefitinib trials. *Journal of Clinical Oncology.* 2005; 23: 8081-8092.
9. Pao W, Miller V, Zakowski M, Doherty J, Politi K, et al. EGF receptor gene mutations are common in lung cancers from "never smokers" and are associated with sensitivity of tumors to gefitinib and erlotinib. *Proc Natl Acad Sci USA.* 2004; 101: 13306-13311.
10. Paez JG, Jänne PA, Lee JC, Tracy S, Greulich H, et al. EGFR mutations in lung cancer: correlation with clinical response to gefitinib therapy. *Science.* 2004; 304: 1497-1500.
11. Lynch TJ, Bell DW, Sordella R, Gurubhagavatula S, Okimoto RA, et al. Activating Mutations in the Epidermal Growth Factor Receptor Underlying Responsiveness of Non-Small-Cell Lung Cancer to Gefitinib. *New England Journal of Medicine.* 2004; 350: 2129-2139.
12. Tsao MS, Sakurada A, Cutz JC, Zhu CQ, Kamel-Reid S, et al. Erlotinib in lung cancer - molecular and clinical predictors of outcome. *N Engl J Med.* 2005; 353: 133-144.
13. US Food and Drug Administration: Drugs@FDA: FDA approved drug products.
14. Shepherd FA, Rodrigues Pereira J, Ciuleanu T, Tan EH, Hirsh V, et al. Erlotinib in Previously Treated Non-Small-Cell Lung Cancer. *New England Journal of Medicine.* 2005; 353: 123-132.
15. Mok TS, Wu YL, Thongprasert S, Yang CH, Chu DT, et al. Gefitinib or carboplatin-paclitaxel in pulmonary adenocarcinoma. *N Engl J Med.* 2009; 361: 947-957.
16. Mitsudomi T, Morita S, Yatabe Y, Negoro S, Okamoto I, et al. Gefitinib versus cisplatin plus docetaxel in patients with non-small-cell lung cancer harbouring mutations of the epidermal growth factor receptor (WJTOG3405): an open label, randomised phase 3 trial. *The Lancet Oncology.* 2010; 11: 121-128.
17. Rosell R, Moran T, Queralt C, Porta R, Cardenal F, et al. Screening for epidermal growth factor receptor mutations in lung cancer. *N Engl J Med.* 2009; 361: 958-967.
18. Zhou C, Wu Y-L, Chen G, Feng J, Liu X-Q, et al. Erlotinib versus chemotherapy as first-line treatment for patients with advanced EGFR mutation-positive non-small-cell lung cancer (OPTIMAL, CTONG-0802): a multicentre, open-label, randomised, phase 3 study. *The Lancet Oncology.* 2011; 12: 735-742.
19. Rosell R, Carcereny E, Gervais R, Vergnenegre A, Massuti B, et al. Erlotinib versus standard chemotherapy as first-line treatment for European patients with advanced EGFR mutation-positive non-small-cell lung cancer (EURTAC): A multicentre, open-label, randomised phase 3 trial. *The Lancet Oncology.* 2012; 13: 239-246.
20. Eberhard DA, Johnson BE, Amler LC, Goddard AD, Heldens SL, et al. Mutations in the epidermal growth factor receptor and in KRAS are predictive and prognostic indicators in patients with non-small-cell lung cancer treated with chemotherapy alone and in combination with erlotinib. *Journal of Clinical Oncology.* 2005; 23: 5900-5909.
21. Jänne PA, Wang X, Socinski MA, Crawford J, Stinchcombe TE, et al. Randomized phase II trial of erlotinib alone or with carboplatin and paclitaxel in patients who were never or light former smokers with advanced lung adenocarcinoma: CALGB 30406 trial. *Journal of Clinical Oncology.* 2012; 30: 2063-2069.
22. Coudert B, Ciuleanu T, Park K, Wu Y-L, Giaccone G, et al. Survival benefit with erlotinib maintenance therapy in patients with advanced Non-Small-Cell Lung Cancer (NSCLC) according to response to first-line chemotherapy. *Annals of Oncology.* 2012; 23: 388-394.
23. Solca F, Dahl G, Zoephel A, Bader G, Sanderson M, et al. Target Binding Properties and Cellular Activity of Afatinib (BIBW 2992), an Irreversible ErbB Family Blocker. *Journal of Pharmacology and Experimental Therapeutics.* 2012; 343: 342-350.
24. Sequist LV, Yang JC-H, Yamamoto N, O'Byrne K, Hirsh V, et al. Phase III Study of Afatinib or Cisplatin Plus Pemetrexed in Patients With Metastatic Lung Adenocarcinoma With EGFR Mutations. *Journal of Clinical Oncology.* 2013; 31: 3327-3334.
25. Köhler J, Schuler M. LUX-Lung 3: redundancy, toxicity or a major step forward? Afatinib as front-line therapy for patients with metastatic EGFR-mutated lung cancer. *Future Oncology.* 2014; 10: 533-540.

26. Wu YL, Zhou C, Hu CP, Feng J, Lu S, et al. Afatinib versus cisplatin plus gemcitabine for first-line treatment of Asian patients with advanced non-small-cell lung cancer harbouring EGFR mutations (LUX-Lung 6): An open-label, randomised phase 3 trial. *The Lancet Oncology*. 2014; 15: 213-222.
27. Yang JCH, Wu YL, Schuler M, Sebastian M, Popat S, et al. Afatinib versus cisplatin-based chemotherapy for EGFR mutation-positive lung adenocarcinoma (LUX-Lung 3 and LUX-Lung 6): Analysis of overall survival data from two randomised, phase 3 trials. *The Lancet Oncology*. 2015; 16: 141-151.
28. Eskens FALM, Mom CH, Planting AST, Gietema JA, Amelsberg A, et al. A phase I dose escalation study of BIBW 2992, an irreversible dual inhibitor of epidermal growth factor receptor 1 (EGFR) and 2 (HER2) tyrosine kinase in a 2-week on, 2-week off schedule in patients with advanced solid tumours. *British Journal of Cancer*. 2008; 98: 80-85.
29. Maemondo M, Inoue A, Kobayashi K, Sugawara S, Oizumi S, et al. Gefitinib or chemotherapy for non-small-cell lung cancer with mutated EGFR. *The New England journal of medicine*. 2010; 362: 2380-2388.
30. Kobayashi S, Boggon TJ, Dayaram T, Jänne PA, Koehler O, et al. EGFR mutation and resistance of non-small-cell lung cancer to gefitinib. *N Engl J Med*. 2005; 352: 786-792.
31. Nguyen K-SH, Kobayashi S, Costa DB. Acquired resistance to epidermal growth factor receptor tyrosine kinase inhibitors in non-small-cell lung cancers dependent on the epidermal growth factor receptor pathway. *Clinical Lung Cancer*. 2009; 10: 281-289.
32. Yu HA, Arcila ME, Rekhtman N, Sima CS, Zakowski MF, et al. Analysis of tumor specimens at the time of acquired resistance to EGFR-TKI therapy in 155 patients with EGFR-mutant lung cancers. *Clinical Cancer Research*. 2013; 19: 2240-2247.
33. Sequist LV, Waltman B, Dias-Santagata D, Digumarthy S, Turke AB, et al. Genotypic and histological evolution of lung cancers acquiring resistance to EGFR inhibitors. *Science Translational Medicine*. 2011; 3: 75ra26.
34. Chong CR, Jänne PA. The quest to overcome resistance to EGFR-targeted therapies in cancer. *Nat Med*. 2013; 19: 1389-1400.
35. Stewart EL, Tan SZ, Liu G, Tsao M. Known and putative mechanisms of resistance to EGFR targeted therapies in NSCLC patients with EGFR mutations - a review. 2015; 4: 67-81.
36. Wilson C, Nicholes K, Bustos D, Lin E, Song Q, et al. Overcoming EMT-associated resistance to anti-cancer drugs via Src/FAK pathway inhibition. *Oncotarget*. 2014; 5: 7328-7341.
37. Han Y, Xiao H, Zhou Z, Yuan M, Zeng Y, et al. Cost-effectiveness analysis of strategies introducing integrated ¹⁸F-FDG PET/CT into the mediastinal lymph node staging of non-small-cell lung cancer. *Nucl Med Commun*. 2015; 36: 234-241.
38. Cross DAE, Ashton SE, Ghiorghiu S, Eberlein C, Nebhan CA, et al. AZD9291, an irreversible EGFR TKI, overcomes T790M-mediated resistance to EGFR inhibitors in lung cancer. *Cancer Discovery*. 2014; 4: 1046-1061.
39. Jänne PA, Yang JC, Kim DW, Planchard D, Ohe Y, et al. AZD9291 in EGFR inhibitor-resistant non-small-cell lung cancer. *N Engl J Med*. 2015; 372: 1689-1699.
40. Yang JCH, Kim DW, Kim SW, Cho BC, Lee JS, et al. Osimertinib activity in patients (pts) with Leptomeningeal (LM) disease from Non-Small Cell Lung Cancer (NSCLC): Updated results from BLOOM, a phase I study. *J Clin Oncol*. 2016; 34.
41. Walter AO, Sjin RTT, Haringsma HJ, Ohashi K, Sun J, et al. Discovery of a Mutant-Selective Covalent Inhibitor of EGFR that Overcomes T790M-Mediated Resistance in NSCLC. *Cancer Discovery*. 2013; 3:1404-1415.
42. Sequist LV, Goldman JW, Wakelee HA, Ross D, Yu HA, et al. Efficacy of Rociletinib (CO-1686) in NSCLC Patients. ASCO Annual Meeting 2015. 2015.
43. Sequist LV, Soria JC, Goldman JW, Wakelee HA, Gadgeel SM, et al. Rociletinib in EGFR-mutated non-small-cell lung cancer. *N Engl J Med*. 2015; 372: 1700-1709.
44. Piotrowska Z, Liu E, Logan J, Chow J, Woreta FA, et al. Rociletinib-associated cataracts in EGFR-mutant NSCLC. *J Clin Oncol*. 2016; 34.
45. Goldman JW, Soria JC, Wakelee HA, Camidge DR, Gadgeel SM, et al. Updated results from TIGER-X, a phase I/II open label study of rociletinib in patients (pts) with advanced, recurrent T790M-positive Non-Small Cell Lung Cancer (NSCLC). *J Clin Oncol*. 2016; 34.
46. Ahn MJ, Kim DW, Kim TM, Lin CC, Ratnayake J, et al. Phase I study of AZD3759, a CNS penetrable EGFR inhibitor, for the treatment of Non-Small-Cell Lung Cancer (NSCLC) with Brain Metastasis (BM) and Leptomeningeal Metastasis (LM). *J Clin Oncol*. 2016; 34.
47. Fenizia F, De Luca A, Pasquale R, Sacco A, Forgione L, et al. EGFR mutations in lung cancer: from tissue testing to liquid biopsy. *Future Oncology*. 2015; 11: 1611-1623.
48. Wilkening S, Hemminki K, Kumar Thirumaran R, Lorenzo Bermejo J, Bonn S, et al. Determination of allele frequency in pooled DNA: comparison of three PCR-based methods. *BioTechniques*. 2005; 39: 853-858.

49. Thress KS, Brant R, Carr TH, Dearden S, Jenkins S, et al. EGFR mutation detection in ctDNA from NSCLC patient plasma: A cross-platform comparison of leading technologies to support the clinical development of AZD9291. *Lung Cancer*. 2015; 90: 509-515.
50. Chiu CH, Yang CT, Shih JY, Huang MS, Su WC, et al. Epidermal Growth Factor Receptor Tyrosine Kinase Inhibitor Treatment Response in Advanced Lung Adenocarcinomas with G719X/L861Q/S768I Mutations. *Journal of Thoracic Oncology*. 2015; 10: 793-799.
51. Xu J, Jin B, Chu T, Dong X, Yang H, et al. EGFR Tyrosine Kinase Inhibitor (TKI) in patients with advanced Non-Small Cell Lung Cancer (NSCLC) harboring uncommon EGFR mutations: A real-world study in China. *Lung Cancer*. 2016; 96: 87-92.
52. Morris SW, Naeve C, Mathew P, James PL, Kirstein MN, et al. ALK, the chromosome 2 gene locus altered by the t(2;5) in non-Hodgkin's lymphoma, encodes a novel neural receptor tyrosine kinase that is highly related to Leukocyte Tyrosine Kinase (LTK). *Oncogene*. 1997;14: 2175-2188.
53. Iwahara T, Fujimoto J, Wen D, Cupples R, Bucay N, et al. Molecular characterization of ALK, a receptor tyrosine kinase expressed specifically in the nervous system. *Oncogene*. 1997; 14: 439-449.
54. Webb TR, Slavish J, George RE, Look AT, Xue L, et al. Anaplastic lymphoma kinase: role in cancer pathogenesis and small-molecule inhibitor development for therapy. *Expert Review of Anticancer Therapy*. 2009; 9: 331-356.
55. Soda M, Choi YL, Enomoto M, Takada S, Yamashita Y, et al. Identification of the transforming EML4-ALK fusion gene in non-small-cell lung cancer. *Nature*. 2007; 448: 561-566.
56. Rikova K, Guo A, Zeng Q, Possemato A, Yu J, et al. Global Survey of Phosphotyrosine Signaling Identifies Oncogenic Kinases in Lung Cancer. *Cell*. 2007; 131: 1190-1203.
57. Blackhall FH, Peters S, Bubendorf L, Dafni U, Kerr KM, et al. Prevalence and clinical outcomes for patients with ALK-positive resected stage I to III adenocarcinoma: Results from the European Thoracic Oncology Platform Lungscape project. *Journal of Clinical Oncology*. 2014; 32: 2780-2787.
58. Takeuchi K, Young LC, Togashi Y, Soda M, Hatano S, et al. KIF5B-ALK, a novel fusion onco kinase identified by an immunohistochemistry- based diagnostic system for ALK-positive lung cancer. *Clinical Cancer Research*. 2009; 15: 3143-3149.
59. Iyevleva AG, Raskin GA, Tiurin VI, Sokolenko AP, Mitiushkina NV, et al. Novel ALK fusion partners in lung cancer. *Cancer Letters*. 2015; 362: 116-121.
60. Choi YL, Takeuchi K, Soda M, Inamura K, Togashi Y, et al. Identification of novel isoforms of the EML4-ALK transforming gene in non-small cell lung cancer. *Cancer Res*. 2008; 68: 4971-4976.
61. Koivunen JP, Mermel C, Zejnullahu K, Murphy C, Lifshits E, et al. EML4-ALK fusion gene and efficacy of an ALK kinase inhibitor in lung cancer. *Clinical Cancer Research*. 2008; 14: 4275-4283.
62. Shinmura K, Kageyama S, Tao H, Bunai T, Suzuki M, et al. EML4-ALK fusion transcripts, but no NPM-, TPM3-, CLTC-, ATIC-, or TFG-ALK fusion transcripts, in non-small cell lung carcinomas. *Lung Cancer*. 2008; 61: 163-169.
63. Inamura K, Takeuchi K, Togashi Y, Hatano S, Ninomiya H, et al. EML4-ALK lung cancers are characterized by rare other mutations, a TTF-1 cell lineage, an acinar histology, and young onset. *Modern Pathology*. 2009; 22: 508-515.
64. Wong DWS, Leung ELH, So KKT, Tam IYS, Sihoe ADL, et al. The EML4-ALK fusion gene is involved in various histologic types of lung cancers from nonsmokers with wild-type EGFR and KRAS. *Cancer*. 2009; 115: 1723-1733.
65. Martelli MP, Sozzi G, Hernandez L, Pettrossi V, Navarro A, et al. EML4-ALK Rearrangement in Non-Small Cell Lung Cancer and Non-Tumor Lung Tissues. *The American Journal of Pathology*. 2009; 174: 661-670.
66. McDermott U, Iafrate AJ, Gray NS, Shioda T, Classon M, et al. Genomic alterations of anaplastic lymphoma kinase may sensitize tumors to anaplastic lymphoma kinase inhibitors. *Cancer Research*. 2008; 68: 3389-3395.
67. Christensen JG, Zou HY, Arango ME, Li Q, Lee JH, et al. Cytoreductive antitumor activity of PF-2341066, a novel inhibitor of anaplastic lymphoma kinase and c-Met, in experimental models of anaplastic large-cell lymphoma. *Molecular Cancer Therapeutics*. 2007; 6: 3314-3322.
68. Kwak EL, Bang YJ, Camidge DR, Shaw AT, Solomon B, et al. Anaplastic Lymphoma Kinase Inhibition in Non-Small-Cell Lung Cancer. *New England Journal of Medicine*. 2010; 363: 1693-1703.
69. Shaw AT, Kim DW, Nakagawa K, Seto T, Crinó L, Ahn MJ, De Pas T. Crizotinib versus chemotherapy in advanced ALK-positive lung cancer. *N Engl J Med*. 2013; 368: 2385-2394.
70. Solomon BJ, Mok T, Kim DW, Wu YL, Nakagawa K, Mekhail T, Felip E. First-line crizotinib versus chemotherapy in ALK-positive lung cancer. *N Engl J Med*. 2014; 371: 2167-2177.

71. Yamaguchi N, Lucena-Araujo AR, Nakayama S, de Figueiredo-Pontes LL, Gonzalez DA, et al. Dual ALK and EGFR inhibition targets a mechanism of acquired resistance to the tyrosine kinase inhibitor crizotinib in ALK rearranged lung cancer. *Lung Cancer*. 2014; 83: 37-43.
72. Doebele RC, Pilling AB, Aisner DL, Kutateladze TG, Le AT, Weickhardt AJ, Kondo KL. Mechanisms of resistance to crizotinib in patients with ALK gene rearranged non-small cell lung cancer. *Clin Cancer Res*. 2012; 18: 1472-1482.
73. Kim S, Kim TM, Kim D-W, Go H, Keam B, et al. Heterogeneity of Genetic Changes Associated with Acquired Crizotinib Resistance in ALK-Rearranged Lung Cancer. *Journal of Thoracic Oncology*. 2013; 8: 415-422.
74. Camidge DR, Pao W, Sequist L V. Acquired resistance to TKIs in solid tumours: learning from lung cancer. *Nature Reviews Clinical Oncology*. 2014; 11: 473-481.
75. Rangachari D, Yamaguchi N, VanderLaan PA, Folch E, Mahadevan A, et al. Brain metastases in patients with EGFR-mutated or ALK-rearranged non-small-cell lung cancers. *Lung Cancer*. 2015; 88: 108-111.
76. Costa DB, Kobayashi S, Pandya SS, Yeo WL, Shen Z, Tan W, Wilner KD. CSF concentration of the anaplastic lymphoma kinase inhibitor crizotinib. *J Clin Oncol*. 2011; 29: e443-445.
77. Costa DB, Shaw AT, Ou SH, Solomon BJ, Riely GJ, et al. Clinical Experience With Crizotinib in Patients With Advanced ALK-Rearranged Non-Small-Cell Lung Cancer and Brain Metastases. *J Clin Oncol*. 2015; 33: 1881-1888.
78. Li N, Michellys P-Y, Kim S, Pferdekamper AC, Li J, et al. Abstract B232: Activity of a potent and selective phase I ALK inhibitor LDK378 in naive and crizotinib-resistant preclinical tumor models. *Molecular Cancer Therapeutics*. 2011; 10: B232-B232.
79. Lovly CM, Shaw AT. Molecular pathways: resistance to kinase inhibitors and implications for therapeutic strategies. *Clin Cancer Res*. 2014; 20: 2249-2256.
80. Katayama R, Shaw AT, Khan TM, Mino-Kenudson M, Solomon BJ, et al. Mechanisms of Acquired Crizotinib Resistance in ALK-Rearranged Lung Cancers. *Science Translational Medicine*. 2012; 4: 120ra17.
81. Kim DW, Mehra R, Tan DS, Felip E, Chow LQ, et al. Activity and safety of ceritinib in patients with ALK-rearranged non-small-cell lung cancer (ASCEND-1): updated results from the multicentre, open-label phase 1 trial. *Lancet Oncol*. 2016; 17: 452-463.
82. Khozin S, Blumenthal GM, Zhang L, Tang S, Brower M, et al. FDA approval: ceritinib for the treatment of metastatic anaplastic lymphoma kinase-positive non-small cell lung cancer. *Clinical cancer research?: an official journal of the American Association for Cancer Research*. 2015; 21: 2436-2439.
83. Shaw AT, Kim DW, Mehra R, Tan DS, Felip E, et al. Ceritinib in ALK-rearranged non-small-cell lung cancer. *N Engl J Med*. 2014; 370: 1189-1197.
84. Sakamoto H, Tsukaguchi T, Hiroshima S, Kodama T, Kobayashi T, et al. CH5424802, a selective ALK inhibitor capable of blocking the resistant gatekeeper mutant. *Cancer Cell*. 2011; 19: 679-690.
85. Kodama T, Tsukaguchi T, Yoshida M, Kondoh O, Sakamoto H. Selective ALK inhibitor alectinib with potent antitumor activity in models of crizotinib resistance. *Cancer Letters*. 2014; 351: 215-221.
86. Seto T, Kiura K, Nishio M, Nakagawa K, Maemondo M, et al. CH5424802 (RO5424802) for patients with ALK-rearranged advanced non-small-cell lung cancer (AF-001JP study): A single-arm, open-label, phase 1-2 study. *The Lancet Oncology*. 2013; 14: 590-598.
87. Gadgeel SM, Gandhi L, Riely GJ, Chiappori AA, West HL, et al. Safety and activity of alectinib against systemic disease and brain metastases in patients with crizotinib-resistant ALK-rearranged non-small-cell lung cancer (AF-002JG): results from the dose-finding portion of a phase 1/2 study. *The Lancet Oncology*. 2014; 15: 1119-1128.
88. Shaw AT, Gandhi L, Gadgeel S, Riely GJ, Cetnar J, et al. Alectinib in ALK-positive, crizotinib-resistant, non-small-cell lung cancer: a single-group, multicentre, phase 2 trial. *Lancet Oncol*. 2016; 17: 234-242.
89. Ou SH, Ahn JS, De Petris L, Govindan R, Yang JC, et al. Alectinib in Crizotinib-Refractory ALK-Rearranged Non-Small-Cell Lung Cancer: A Phase II Global Study. *J Clin Oncol*. 2016; 34: 661-668.
90. Nokihara H, Hida T, Kondo M, Kim YH, Azuma K, et al. Alectinib (ALC) versus crizotinib (CRZ) in ALK-inhibitor naive ALK-positive non-small cell lung cancer (ALK+ NSCLC): Primary results from the J-ALEX study. *J Clin Oncol*. 2016; 34.
91. Zou HY, Li Q, Engstrom LD, West M, Appleman V, et al. PF-06463922 is a potent and selective next-generation ROS1/ALK inhibitor capable of blocking crizotinib-resistant ROS1 mutations. *Proc Natl Acad Sci U S A*. 2015; 112: 3493-3498.
92. Solomon BJ, Bauer TM, Felip E, Besse B, James LP, et al. Safety and efficacy of lorlatinib (PF-06463922) from the dose-escalation component of a study in patients with advanced ALK+ or ROS1+ Non-Small Cell Lung Cancer (NSCLC). *J Clin Oncol*. 2016; 34.
93. Bauer T, Solomon B, Besse B, Navarro A, James L, et al. Clinical Activity and Safety of the ALK/ROS1 TK Inhibitor PF-06463922 in Advanced NSCLC. In: *WCLC*. Denver. 2015. Abstract ORAL 33.07. 2015.

94. Shaw AT, Friboulet L, Leshchiner I, Gainor JF, Bergqvist S, et al. Resensitization to Crizotinib by the Lorlatinib ALK Resistance Mutation L1198F. *N Engl J Med*. 2016; 374: 54-61.
95. Huang WS, Liu S, Zou D, Thomas M, Wang Y, et al. Discovery of Brigatinib (AP26113), a Phosphine Oxide-Containing, Potent, Orally Active Inhibitor of Anaplastic Lymphoma Kinase. *Journal of Medicinal Chemistry*. 2016.
96. Zhang S, Nadworny S, Wardwell SD, Eichinger L, Das B, et al. Abstract 781: The potent ALK inhibitor AP26113 can overcome mechanisms of resistance to first- and second-generation ALK TKIs in preclinical models. *Cancer Research*. 2015; 75: 781-781.
97. Rosell R, Gettinger SN, Bazhenova LA, Langer CJ, Salgia R, et al. 1330: Brigatinib efficacy and safety in patients (Pts) with anaplastic lymphoma kinase (ALK)-positive (ALK+) non-small cell lung cancer (NSCLC) in a phase 1/2 trial. *Journal of Thoracic Oncology*. 2016; 11: 113-142.
98. Kim DW, Tiseo M, Ahn MJ, Reckamp KL, Hansen KH, et al. Brigatinib (BRG) in patients (pts) with crizotinib (CRZ)-refractory ALK+ non-small cell lung cancer (NSCLC): First report of efficacy and safety from a pivotal randomized phase (ph) 2 trial (ALTA). *J Clin Oncol*. 2016; 34.
99. Acquaviva J, Wong R, Charest A. The multifaceted roles of the receptor tyrosine kinase ROS in development and cancer. *Biochimica et Biophysica Acta (BBA) - Reviews on Cancer*. 2009; 1795: 37-52.
100. Robinson DR, Wu YM, Lin SF. The protein tyrosine kinase family of the human genome. *Oncogene*. 2000; 19: 5548-5557.
101. Gainor JF, Shaw AT. Novel Targets in Non-Small Cell Lung Cancer: ROS1 and RET Fusions. *The Oncologist*. 2013; 18: 865-875.
102. Bergethon K, Shaw AT, Ou SHI, Katayama R, Lovly CM, et al. ROS1 rearrangements define a unique molecular class of lung cancers. *Journal of Clinical Oncology*. 2012; 30: 863-870.
103. Rimkunas VM, Crosby KE, Li D, Hu Y, Kelly ME, et al. Analysis of Receptor Tyrosine Kinase ROS1-Positive Tumors in Non-Small Cell Lung Cancer: Identification of a FIG-ROS1 Fusion. *Clinical Cancer Research*. 2012; 18: 4449-4457.
104. Shaw AT, Ou SH, Bang YJ, Camidge DR, Solomon BJ, et al. Crizotinib in ROS1-rearranged non-small-cell lung cancer. *N Engl J Med*. 2014; 371: 1963-1971.
105. Awad MM, Katayama R, McTigue M, Liu W, Deng YL, Brooun A, Friboulet L. Acquired resistance to crizotinib from a mutation in CD74-ROS1. *N Engl J Med*. 2013; 368: 2395-2401.
106. Dilon A, Somwar R, Wagner JP, Vellore NA, Eide A, et al. A novel crizotinib-resistant solvent-front mutation responsive to cabozantinib therapy in a patient with ROS1-rearranged lung cancer. *Clinical Cancer Research*. 2015; 1: OF1-OF8.
107. Shaw AT, Bauer TM, Felip E, Besse B, James LP, et al. Clinical activity and safety of PF-06463922 from a dose escalation study in patients with advanced ALK+ or ROS1+ NSCLC. *J Clin Oncol*. 2015; 33.
108. Katayama R, Kobayashi Y, Friboulet L, Lockerman EL, Koike S, et al. Cabozantinib Overcomes Crizotinib Resistance in ROS1 Fusion-Positive Cancer. *Clinical Cancer Research*. 2015; 21: 166-174.
109. Slebos RJ, Kibbelaar RE, Dalesio O, Kooistra A, Stam J, et al. K-ras oncogene activation as a prognostic marker in adenocarcinoma of the lung. *N Engl J Med*. 1990; 323: 561-565.
110. Kris MG, Johnson BE, Berry LD, Kwiatkowski DJ, Iafrate AJ, et al. Using multiplexed assays of oncogenic drivers in lung cancers to select targeted drugs. *JAMA*. 2014; 311: 1998-2006.
111. Wood K, Hensing T, Malik R, Salgia R. Prognostic and Predictive Value in KRAS in Non-Small-Cell Lung Cancer. *JAMA Oncology*. 2016; 91010.
112. Ahrendt SA, Decker PA, Alawi EA, Zhu Yr YR, Sanchez-Cespedes M, et al. Cigarette smoking is strongly associated with mutation of the K-ras gene in patients with primary adenocarcinoma of the lung. *Cancer*. 2001; 92: 1525-1530.
113. Graziano SL, Gamble GP, Newman NB, Abbott LZ, Rooney M, et al. Prognostic significance of K-ras codon 12 mutations in patients with resected stage I and II non-small-cell lung cancer. *J Clin Oncol*. 1999; 17: 668-675.
114. Broermann P, Junker K, Brandt BH, Heinecke A, Freitag L, et al. Trimodality treatment in Stage III nonsmall cell lung carcinoma: prognostic impact of K-ras mutations after neoadjuvant therapy. *Cancer*. 2002; 94: 2055-2062.
115. Grossi F, Loprevite M, Chiaramondia M, Ceppa P, Pera C, et al. Prognostic significance of K-ras, p53, bcl-2, PCNA, CD34 in radically resected non-small cell lung cancers. *European Journal of Cancer*. 2003; 39: 1242-1250.
116. Schneider C-P, Heigener D, Schott-von-Römer K, Gütz S, Laack E, et al. Epidermal Growth Factor Receptor-Related Tumor Markers and Clinical Outcomes with Erlotinib in Non-small Cell Lung Cancer: An Analysis of Patients from German Centers in the TRUST Study. *Journal of Thoracic Oncology*. 2008; 3: 1446-1453.
117. Brugger W, Triller N, Blasinska-Morawiec M, Curescu S, Sakalauskas R, et al. Prospective molecular marker analyses of EGFR and KRAS from a randomized, placebo-controlled study of erlotinib maintenance therapy in advanced non-small-cell lung cancer. *Journal of Clinical Oncology*. 2011; 29: 4113-4120.

118. Stinchcombe TE, Johnson GL. MEK inhibition in non-small cell lung cancer. *Lung Cancer*. 2014; 86: 121-125.
119. Jänne PA, Shaw AT, Pereira JR, Jeannin G, Vansteenkiste J, et al. Selumetinib plus docetaxel for KRAS-mutant advanced non-small-cell lung cancer: a randomised, multicentre, placebo-controlled, phase 2 study. *Lancet Oncology*. 2013; 14: 38-47.
120. Carter CA, Rajan A, Keen C, Szabo E, Khozin S, et al. Selumetinib with and without erlotinib in KRAS mutant and KRAS wild-type advanced nonsmall-cell lung cancer. *Annals of Oncology*. 2016; 27: 693-699.
121. Roberts PJ, Stinchcombe TE. KRAS mutation: Should we test for it, and does it matter. *Journal of Clinical Oncology*. 2013; 31: 1112-1121.
122. Liu L, Hong S, Zhang V, Gilmer T. Abstract 4394: Identification of molecular determinants of response to GSK1120212B, a potent and selective MEK inhibitor, as a single agent and in combination in RAS/RAF mutant non-small cell lung carcinoma cells. *Cancer Research*. 2011; 71: 4394-4394.
123. Gandara DR, Hirt S, Blumenschein GR, Barlesi F, Delord J-P, et al. Oral MEK1/MEK2 inhibitor trametinib (GSK1120212) in combination with docetaxel in KRAS-mutant and Wild-Type (WT) advanced Non-Small Cell Lung Cancer (NSCLC): A phase I/Ib trial. *ASCO Meeting Abstracts*. 2013; 31: 8028.
124. Sithanandam G, Druck T, Cannizzaro LA, Leuzzi G, Huebner K, et al. B-raf and a B-raf pseudogene are located on 7q in man. *Oncogene*. 1992; 7: 795-799.
125. Paik PK, Arcila ME, Fara M, Sima CS, Miller VA, et al. Clinical characteristics of patients with lung adenocarcinomas harboring BRAF mutations. *J Clin Oncol*. 2011; 29: 2046-2051.
126. Marchetti A, Felicioni L, Malatesta S, Grazia Sciarrotta M, Guetti L, et al. Clinical features and outcome of patients with non-small-cell lung cancer harboring BRAF mutations. *J Clin Oncol*. 2011; 29: 3574-3579.
127. Gautschi O, Pauli C, Strobel K, Hirschmann A, Printzen G, et al. A Patient with BRAF V600E Lung Adenocarcinoma Responding to Vemurafenib. *Journal of Thoracic Oncology*. 2012; 7: e23-e24.
128. Peters S, Michielin O, Zimmermann S. Dramatic response induced by vemurafenib in a BRAF V600E-mutated lung adenocarcinoma. *Journal of Clinical Oncology*. 2013; 31.
129. Robinson SD, O'Shaughnessy JA, Lance Cowey C, Konduri K. BRAF V600E-mutated lung adenocarcinoma with metastases to the brain responding to treatment with vemurafenib. *Lung Cancer*. 2014; 85: 326-330.
130. Planchard D, Julien Mazieres, Gregory J Riely, Charles M Rudin, Fabrice Barlesi, et al. Interim results of phase II study BRF113928 of dabrafenib in BRAF V600E mutation-positive non-small cell lung cancer patients. *ASCO Abstract 8009*. 2013.
131. Hyman DM, Puzanov I, Subbiah V, Faris JE, Chau I, et al. Vemurafenib in Multiple Non melanoma Cancers with BRAF V600 Mutations. *N Engl J Med*. 2015; 373: 726-736.
132. Planchard D, Groen HJM, Kim TM, Rigas JR, Souquet PJ, et al. Interim results of a phase II study of the BRAF inhibitor (BRAFi) dabrafenib (D) in combination with the MEK inhibitor trametinib (T) in patients (pts) with BRAF V600E mutated (mut) metastatic Non-Small Cell Lung Cancer (NSCLC). *J Clin Oncol*. 2015; 33.
133. Ju YS, Lee WC, Shin JY, Lee S, Bleazard T, et al. A transforming KIF5B and RET gene fusion in lung adenocarcinoma revealed from whole-genome and transcriptome sequencing. *Genome Research*. 2012; 22: 436-445.
134. Wang R, Hu H, Pan Y, Li Y, Ye T, et al. RET fusions define a unique molecular and clinicopathologic subtype of non-small-cell lung cancer. *Journal of Clinical Oncology*. 2012; 30: 4352-4359.
135. Chao BH, Briesewitz R, Villalona-Calero MA. RET Fusion Genes in Non-Small-Cell Lung Cancer. *Journal of Clinical Oncology*. 2012; 30: 4439-4441.
136. Drlon A, Wang L, Hasanovic A, Suehara Y, Lipson D, et al. Response to cabozantinib in patients with RET fusion-positive lung adenocarcinomas. *Cancer Discovery*. 2013; 3: 630-635.
137. Lipson D, Capelletti M, Yelensky R, Otto G, Parker A, et al. Identification of new ALK and RET gene fusions from colorectal and lung cancer biopsies. *Nat Med*. 2012; 18: 382-384.
138. Kohno T, Ichikawa H, Totoki Y, Yasuda K, Hiramoto M, et al. KIF5B-RET fusions in lung adenocarcinoma. *Nature Medicine*. 2012; 18: 375-377.
139. Takeuchi K, Soda M, Togashi Y, Suzuki R, Sakata S, et al. RET, ROS1 and ALK fusions in lung cancer. *Nature medicine*. 2012; 18: 378-381.
140. Okamoto K, Kodama K, Takase K, Sugi NH, Yamamoto Y, et al. Antitumor activities of the targeted multi-tyrosine kinase inhibitor lenvatinib (E7080) against RET gene fusion-driven tumor models. *Cancer Letters*. 2013; 340: 97-103.
141. Socinski MA, Novello S, Brahmer JR, Rosell R, Sanchez JM, et al. Multicenter, phase II trial of sunitinib in previously treated, advanced non-small-cell lung cancer. *Journal of Clinical Oncology*. 2008; 26: 650-656.

142. Blumenschein GR, Gatzemeier U, Fossella F, Stewart DJ, Cupit L, et al. Phase II, multicenter, uncontrolled trial of single-agent sorafenib in patients with relapsed or refractory, advanced non-small-cell lung cancer. *Journal of Clinical Oncology*. 2009; 27: 4274-4280.
143. Lee JS, Hirsh V, Park K, Qin S, Blajman CR, et al. Vandetanib Versus Placebo in Patients With Advanced Non-Small-Cell Lung Cancer After Prior Therapy With an Epidermal Growth Factor Receptor Tyrosine Kinase Inhibitor: A Randomized, Double-Blind Phase III Trial (ZEPHYR). *Journal of Clinical Oncology*. 2012; 30: 1114-1121.
144. Gautschi O, Zander T, Keller FA, Strobel K, Hirschmann A, et al. A Patient with Lung Adenocarcinoma and RET Fusion Treated with Vandetanib. *Journal of Thoracic Oncology*. 2013; 8: e43-e44.
145. Falchook GS, Ordóñez NG, Bastida CC, Stephens PJ, Miller VA, et al. Effect of the RET Inhibitor Vandetanib in a Patient with RET Fusion-Positive Metastatic Non-Small-Cell Lung Cancer. *J Clin Oncol*. 2016; 34: e141-144.
146. Gautschi O, Wolf J, Milia J, Filleron T, Carbone D, et al. Targeting RET in patients with RET-rearranged lung cancers: results from a global registry. ASCO 2016. Abstract. 2014.
147. Mazieres J, Peters S, Lepage B, Cortot AB, Barlesi F, et al. Lung Cancer That Harbors an HER2 Mutation: Epidemiologic Characteristics and Therapeutic Perspectives. *Journal of Clinical Oncology*. 2013; 31: 1997-2003.
148. Buttiatta F, Barassi F, Fresu G, Felicioni L, Chella A, et al. Mutational analysis of the HER2 gene in lung tumors from Caucasian patients: Mutations are mainly present in adenocarcinomas with bronchioloalveolar features. *International Journal of Cancer*. 2006; 119: 2586-2591.
149. Perera SA, Li D, Shimamura T, Raso MG, Ji H, et al. HER2YVMA drives rapid development of adenosquamous lung tumors in mice that are sensitive to BIBW2992 and rapamycin combination therapy. *Proceedings of the National Academy of Sciences*. 2009; 106: 474-479.
150. Heinmöller P, Gross C, Beyser K, Schmidtgen C, Maass G, et al. HER2 status in non-small cell lung cancer: results from patient screening for enrollment to a phase II study of herceptin. *Clinical cancer research? An official journal of the American Association for Cancer Research*. 2003; 9: 5238-5243.
151. Stephens P, Hunter C, Bignell G, Edkins S, Davies H, et al. Lung cancer: intragenic ERBB2 kinase mutations in tumours. *Nature*. 2004; 431: 525-526.
152. Li C, Fang R, Sun Y, Han X, Li F, et al. Spectrum of oncogenic driver mutations in lung adenocarcinomas from East Asian never smokers. *PLoS ONE*. 2011; 6.
153. Langer CJ, Stephenson P, Thor A, Vangel M, Johnson DH. Trastuzumab in the treatment of advanced non-small-cell lung cancer: Is there a role? Focus on Eastern Cooperative Oncology Group Study 2598. *Journal of Clinical Oncology*. 2004; 22: 1180-1187.
154. Gatzemeier U, Groth G, Butts C, Van Zandwijk N, Shepherd F, Arzidzoni A, Barton C. Randomized phase II trial of gemcitabine-cisplatin with or without trastuzumab in HER2-positive non-small-cell lung cancer. *Ann Oncol*. 2004; 15: 19-27.
155. Hirsch FR, Langer CJ. The role of HER2/neu expression and trastuzumab in non-small cell lung cancer. *Seminars in oncology*. 2004; 31: 75-82.
156. De Grève J, Teugels E, Geers C, Decoster L, Galdermans D, et al. Clinical activity of afatinib (BIBW 2992) in patients with lung adenocarcinoma with mutations in the kinase domain of HER2/neu. *Lung Cancer*. 2012; 76: 123-127.
157. Kris MG, Camidge DR, Giaccone G, Hida T, Li BT, et al. Targeting HER2 aberrations as actionable drivers in lung cancers: Phase II trial of the pan-HER tyrosine kinase inhibitor dacomitinib in patients with HER2-mutant or amplified tumors. *Annals of Oncology*. 2015; 26: 1421-1427.
158. Seki T, Hagiya M, Shimonishi M, Nakamura T, Shimizu S. Organization of the human hepatocyte growth factor-encoding gene. *Gene*. 1991; 102: 213-219.
159. Engelman JA, Zejnullahu K, Mitsudomi T, Song Y, Hyland C, et al. MET amplification leads to gefitinib resistance in lung cancer by activating ERBB3 signaling. *Science*. 2007; 316: 1039-1043.
160. Beau-Faller M, Ruppert AM, Voegelé AC, Neuville A, Meyer N, et al. MET gene copy number in non-small cell lung cancer: molecular analysis in a targeted tyrosine kinase inhibitor naïve cohort. *J Thorac Oncol*. 2008; 3: 331-339.
161. Park S, Choi Y, La, Sung CO, An J, Seo J, et al. High MET copy number and MET over expression: Poor outcome in non-small cell lung cancer patients. *Histology and Histopathology*. 2012; 27: 197-207.
162. Bean J, Brennan C, Shih JY, Riely G, Viale A, et al. MET amplification occurs with or without T790M mutations in EGFR mutant lung tumors with acquired resistance to gefitinib or erlotinib. *Proceedings of the National Academy of Sciences of the United States of America*. 2007; 104: 20932-20937.
163. Onozato R, Kosaka T, Kuwano H, Sekido Y, Yatabe Y, et al. Activation of MET by gene amplification or by splice mutations deleting the juxtamembrane domain in primary resected lung cancers. *Journal of Thoracic Oncology*. 2009; 4: 5-11.

164. Camidge DR, Ou S-HI, Shapiro G, Otterson GA, Villaruz LC, et al. Efficacy and safety of crizotinib in patients with advanced c-MET-amplified Non-Small Cell Lung Cancer (NSCLC). ASCO Meeting Abstracts. 2014; 35: 8001.
165. Vassal G. Activity of crizotinib in relapsed MET amplified malignancies: Results of the French AcS Program. ASCO Meeting Abstracts. 2015; 33: Abstract 2595.
166. Scagliotti G, Novello S, Ramlau R, Favaretto A, Barlesi F, et al. Results of the Phase 3 MARQUEE Study?: MET Inhibitor Tivantinib (ARQ 197) Plus. European Cancer Congress. 2013; (Arq 197).
167. Ou S-HI, Govindan R, Eaton KD, Otterson GA, Gutierrez M, et al. Phase I/II dose-finding study of crizotinib (CRIZ) in combination with erlotinib (E) in patients (pts) with advanced Non-Small Cell Lung cancer (NSCLC). ASCO Meeting Abstracts. 2012; 30: 2610.
168. Wakelee HA, Gettinger SN, Engelman JA, Janne PA, West HJ, et al. A phase Ib/II study of XL184 (BMS 907351) with and without erlotinib (E) in patients (pts) with Non-Small Cell Lung Cancer (NSCLC). ASCO Meeting Abstracts. 2010; 28: 3017.
169. Goldman JW, Laux I, Chai F, Savage RE, Ferrari D, et al. Phase 1 dose-escalation trial evaluating the combination of the selective MET (mesenchymal-epithelial transition factor) inhibitor tivantinib (ARQ 197) plus erlotinib. Cancer. 2012; 118: 5903-5911.
170. Heist RS, Shim HS, Gingipally S, Mino-Kenudson M, Le L, et al. MET Exon 14 Skipping in Non-Small Cell Lung Cancer. The Oncologist. 2016; 21: 481-486.
171. Paik PK, Drilon A, Fan P-D, Yu H, Rekhtman N, et al. Response to MET Inhibitors in Patients with Stage IV Lung Adenocarcinomas Harboring MET Mutations Causing Exon 14 Skipping. Cancer discovery. 2015; 5: 842-849.
172. Stambolic V, Suzuki A, de la Pompa JL, Brothers GM, Mirtsos C, et al. Negative Regulation of PKB/Akt-Dependent Cell Survival by the Tumor Suppressor PTEN. Cell. 1998; 95: 29-39.
173. Zhou BP, Liao Y, Xia W, Spohn B, Lee MH, et al. Cytoplasmic localization of p21Cip1/WAF1 by Akt-induced phosphorylation in HER-2/neu-over expressing cells. Nature cell biology. 2001; 3: 245-252.
174. Vivanco I, Sawyers CL. The phosphatidylinositol 3-Kinase AKT pathway in human cancer. Nat Rev Cancer. 2002; 2: 489-501.
175. Tang JM, He QY, Guo RX, Chang XJ. Phosphorylated Akt overexpression and loss of PTEN expression in non-small cell lung cancer confers poor prognosis. Lung Cancer. 2006; 51: 181-191.
176. Papadimitrakopoulou V. Development of PI3K/AKT/mTOR Pathway Inhibitors and Their Application in Personalized Therapy for Non-Small-Cell Lung Cancer. Journal of Thoracic Oncology. 2012; 7: 1315-1326.
177. Wang L, Hu H, Pan Y, Wang R, Li Y, et al. PIK3CA mutations frequently coexist with EGFR/KRAS mutations in non-small cell lung cancer and suggest poor prognosis in EGFR/KRAS wildtype subgroup. PLoS One. 2014; 9: e88291.
178. Bianco R, Shin I, Ritter CA, Yakes FM, Basso A, et al. Loss of PTEN/MMAC1/TEP in EGF receptor-expressing tumor cells counteracts the antitumor action of EGFR tyrosine kinase inhibitors. Oncogene. 2003; 22: 2812-2822.
179. Gadgeel SM, Wozniak A. Preclinical Rationale for PI3K/Akt/mTOR Pathway Inhibitors as Therapy for Epidermal Growth Factor Receptor Inhibitor-Resistant Non-Small-Cell Lung Cancer. Clinical Lung Cancer. 2013; 14: 322-332.
180. Shapiro GI. Clinical combination of the MEK inhibitor GDC-0973 and the pan-PI3K inhibitor GDC-0941: A first-in-human phase 1b study testing daily and intermittent dosing schedules in patients with advanced solid tumors. ASCO. 2011: Abstract 3005.
181. Besse B, Soria J, Ware J, Adjei A, Dy G, et al. A phase Ib study to evaluate the PI3-kinase inhibitor GDC-0941 with paclitaxel (P) and carboplatin (C), with and without bevacizumab (BEV), in patients with advanced Non-Small Cell Lung Cancer (NSCLC). Journal of clinical oncology, 2011 ASCO Annual Meeting Abstracts Part 1. 2011; 29: 3044.
182. Reungwetwattana T, Molina JR, Mandrekar SJ, Allen-Ziegler K, Rowland KM, et al. Brief Report: A Phase II "Window-of-Opportunity" Frontline Study of the mTOR Inhibitor, Temsirolimus Given as a Single Agent in Patients with Advanced NSCLC, an NCCTG Study. Journal of Thoracic Oncology. 2012; 7: 919-922.
183. Besse B, Leigh N, Bannoun J, Papadimitrakopoulou VA, Blais N, et al. Phase II study of everolimus-erlotinib in previously treated patients with advanced non-small-cell lung cancer. Annals of Oncology. 2014; 25: 409-415.
184. Eberhardt WEE, Mitchell P, Schiller JH, Brown MP, Thomas M, et al. Feasibility of adding everolimus to carboplatin and paclitaxel, with or without bevacizumab, for treatment-naive, advanced non-small cell lung cancer. Investigational New Drugs. 2014; 32: 123-134.
185. Ramalingam SS, Owonikoko TK, Behera M, Subramanian J, Saba NF, et al. Phase II Study of Docetaxel in Combination with Everolimus for Second- or Third-Line Therapy of Advanced Non-Small-Cell Lung Cancer. Journal of Thoracic Oncology. 2013; 8: 369-372.
186. Cheng H, Shcherba M, Pendurti G, Liang Y, Piperdi B, et al. Targeting the PI3K/AKT/mTOR pathway: potential for lung cancer treatment. Lung Cancer Manag. 2014; 3: 67-75.
187. Cheng H, Zou Y, Ross JS, Wang K, Liu X, et al. RICTOR Amplification Defines a Novel Subset of Patients with Lung Cancer Who May Benefit from Treatment with mTORC1/2 Inhibitors. Cancer Discovery. 2015; 5: 1262-1270.

188. Politi K, Herbst RS. Lung cancer in the era of precision medicine. *Clin Cancer Res.* 2015; 21: 2213-2220.
189. Gandara DR, Hammerman PS, Sos ML, Lara PN, Hirsch FR. Squamous Cell Lung Cancer: From Tumor Genomics to Cancer Therapeutics. *Clinical Cancer Research.* 2015; 21: 2236-2243.
190. Clark GM, Zborowski DM, Santabarbara P, Ding K, Whitehead M, et al. Smoking history and epidermal growth factor receptor expression as predictors of survival benefit from erlotinib for patients with non-small-cell lung cancer in the National Cancer Institute of Canada Clinical Trials Group study BR.21. *Clinical lung cancer.* 2006; 7: 389-394.
191. Wojtowicz-Praga S LL. Comparative Efficacy and Safety of Erlotinib in Non-Small Cell Lung Cancer of Squamous Cell and Adenocarcinoma Histology in the Phase 3 NCIC CTG BR.21 and SATURN (BO18192) Trials. *Ann Oncol.* 2012; 23: ix419. abstr.
192. Soria JC, Felip E, Cobo M, Lu S, Syrigos K, et al. Afatinib versus erlotinib as second-line treatment of patients with advanced squamous cell carcinoma of the lung (LUX-Lung 8): an open-label randomised controlled phase 3 trial. *Lancet Oncol.* 2015; 16: 897-907.
193. Kuenen B, Witteveen PO, Ruijter R, Giaccone G, Dontabhaktuni A, et al. A phase I pharmacologic study of necitumumab (IMC-11F8), a fully human IgG1 monoclonal antibody directed against EGFR in patients with advanced solid malignancies. *Clinical Cancer Research.* 2010; 16: 1915-1923.
194. Thatcher N, Hirsch FR, Luft AV, Szczesna A, Ciuleanu TE, et al. Necitumumab plus gemcitabine and cisplatin versus gemcitabine and cisplatin alone as first-line therapy in patients with stage IV squamous non-small-cell lung cancer (SQUIRE): an open-label, randomised, controlled phase 3 trial. *The Lancet. Oncology.* 2015; 16: 763-774.